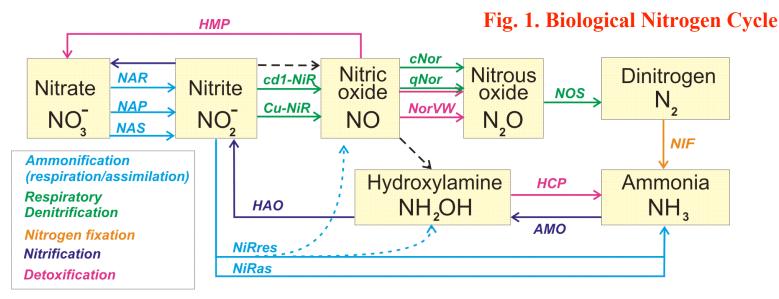
Group of Subsystems: Nitrogen oxides metabolism

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Nitrogen is an essential element in all living organisms. Inter-conversions of nitrogen species between a number of redox states (+5 to -3) form the biogeochemical nitrogen cycle which has multiple environmental impacts and industrial applications [1]. Inorganic nitrogen oxides should be first reduced to ammonium, which can be further incorporated into organic matter via glutamine synthase. Diazotrophic prokaryota possess nitrogenase genes and are able to fix molecular nitrogen from the atmosphere.

On the other hand, bacteria can obtain metabolic energy by redox processes utilizing soluble nitrogen oxides, nitrate and nitrite as terminal respiratory oxidants under oxygen limiting conditions. Two dissimilar pathways of nitrate respiration, ammonification and denitrification, involve formation of a common intermediate, i.e. nitrite, but end in different products, ammonia and gaseous nitrogen oxides (NO or N_2O) or dinitrogen respectively (Fig. 1).

Autotrophic nitrification is a two-step process of an oxidative conversion of ammonia to nitrite via hydroxylamine, carried out by ammonia-oxidizing bacteria, and further oxidation of nitrite to nitrate, performed by nitrite-oxidazing chemolithoautotrophic bacteria.

Finally, the cell should be able to detoxify the exogenously/metabolically produced NO and reactive nitrogen species.

Subsystem: Nitrate and Nitrite ammonification

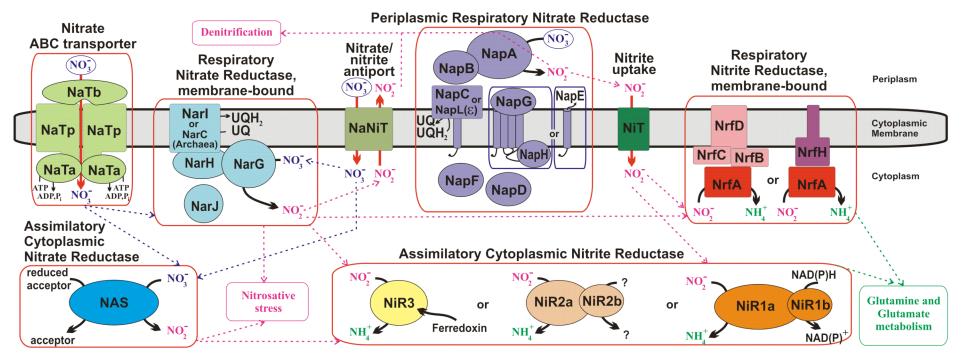
In the first step of this pathway nitrite is formed by one of the three different types of nitrate reductases: soluble assimilatory NAS, membrane-associated respiratory NAR, and periplasmic dissimilatory NAP [2, 3]. NAS is located in the cytoplasmic compartment and participates in nitrogen assimilation (termed NaRas here). NAR is usually a three-subunit complex anchored at the cytoplasmic face of the membrane with its active site located in the cytoplasmic compartment. It is involved in anaerobic nitrate respiration. NAP is a two-subunit complex located in the periplasmic compartment. It is coupled to quinol oxidation via a membrane anchored tetraheme cytochrome.

The members of all three classes of enzymes bind a bis-molybdopterin guanine dinucleotide cofactor at their active sites, but they differ markedly in the number and nature of cofactors used to transfer electrons to this site. Analysis of prokaryotic genomes reveals that different nitrate reductases are phylogenetically widespread.

The next step of ammonification is conversion of nitrite into ammonia by either membrane-bound cytochrome *c* containing respiratory nitrite reductase NrfA, or by one of the three different cytoplasmic assimilatory NiR isoenzymes. In ε - and δ -proteobacteria NrfA forms a stable complex with a transmembrane component NrfH, whereas in γ -proteobacteria NrfH is thought to be replaced by the *nrfBCD* gene products [4]. Among soluble NiRs the siroheme-containing NADPH-dependent enzyme (NirBD in *E.coli*) is the most common one. Cyanobacteria, plants, and some α -proteobacteria possess a distinct ferredoxin-dependent cytoplasmic NiR [5]. Some strictly anaerobic species (e.g. *Clostridia*) have another two-component NiR, which has not yet been characterized [6].

The topological arrangements of nitrate and nitrite reductases in bacteria necessitate synthesis of transporter proteins that carry nitrogen oxyanions across the cytoplasmic membrane. Two types of uptake systems are known to act in assimilation of nitrate (and nitrite): (i) ATP hydrolysis driven ABC transporters, and (ii) secondary transporters reliant on proton motive force, which belong to either nitrite/nitrate transporter family (NarK), nitrite uptake NirC family, or formate/nitrite transporter family [7].

Fig. 2. Nitrate and Nitrite ammonification. Subsystem diagram.



	#	Abbrev.	Functional Role	Subset	#	Abbrev.	Functional Role	Subset
	1	NrfA	Cytochrome c552 precursor (EC 1.7.2.2)		17	NAS	Assimilatory nitrate reductase large subunit (EC:1.7.99.4)	NAS
	2	NrfB	Cytochrome c-type protein nrfB precursor	*NiRres	18	NarG	Respiratory nitrate reductase alpha chain (EC 1.7.99.4)	
	3	NrfC	NrfC protein		19	NarH	Respiratory nitrate reductase beta chain (EC 1.7.99.4)	
	4	NrfD	NrfD protein		20	Narl	Respiratory nitrate reductase gamma chain (EC 1.7.99.4)	*NAR
	5	NrfH	Cytochrome c nitrite reductase, small subunit NrfH		21	NarJ	Respiratory nitrate reductase delta chain (EC 1.7.99.4)	
	6	NiR1a	Nitrite reductase [NAD(P)H] large subunit (EC 1.7.1.4)			NarC	Respiratory nitrate reductase subunit, conjectural (EC 1.7.99.4)	
	7	NiR1b	Nitrite reductase [NAD(P)H] small subunit (EC 1.7.1.4)		23 24	NapA	Periplasmic nitrate reductase precursor (EC 1.7.99.4) Nitrate reductase cytochrome c550-type subunit	
	8	NiR2a	Nitrite reductase probable [NAD(P)H] subunit (EC 1.7.1.4)	*NiRas	24	NapB NapC	Cytochrome c-type protein NapC	
	-	NiR2b	Nitrite reductase probable electron transfer 4Fe-S subunit (EC	MINAS	26	NapC	Periplasmic nitrate reductase component NapD	
_	_		•		27	NapE	Ferredoxin-type protein NapF (periplasmic nitrate reductase)	*NAP
-	-	NiR3	Ferredoxinnitrite reductase (EC 1.7.7.1)	*****	28	NapG	Ferredoxin-type protein NapG (periplasmic nitrate reductase)	
-	-	NiTa	Nitrite transporter from formate/nitrite family	*NiT	29	NapH	Polyferredoxin NapH (periplasmic nitrate reductase)	I I
_1	2	NiTc	Nitrite transporter NirC		30	NapE	Periplasmic nitrate reductase component NapE	I I
1	3	NaTb	Nitrate ABC transporter, nitrate-binding protein	*NaT	31	NapL	Periplasmic nitrate reductase component NapL	
1	4	NaTp	Nitrate ABC transporter, permease protein		32	NaNiRR	Nitrate/nitrite response regulator protein	
1	5	NaTa	Nitrate ABC transporter, ATP-binding protein		33	NaNiS	Nitrate/nitrite sensor protein (EC 2.7.3)	*Reg
1	6	NaNiT	Nitrate/nitrite transporter	NaNiT	34	NasT	Response regulator NasT	1

Fig. 3. Nitrate and Nitrite ammonification. Subsystem spreadsheet.

Organism	Variant Code	NiR respiratory		NiR assimilatory		NaR assimilatory	NaR respiratory	NaR periplasmic	Nitrite and Nitrate transporters			
		NrfA	NrfBCD or NrfH	NiR1a-6, NiR2a-8, NiR3-10	NiR1b- 7, NiR2b-9	NAS	NarGHIJ	NapABCDFGH or NapABCDFE	NiTa-11, NiTc-12	NaTbpa	NaNiT	
Escherichia coli K12	1	<u>3980</u>	NrfBCD	<u>3300</u> -6	<u>3301</u> -7		NarGHIJ	NapABCDFGH	<u>3302</u> -12		<u>1455, 1212</u>	
Bacillus subtilis	2			<u>331</u> -6, <u>333</u> -6	<u>330</u> -7	<u>332</u>	NarGHIJ				<u>334, 3739</u>	
Vibrio vulnificus CMCP6	3	<u>2794</u>	NrfBCD	<u>3318</u> -6, 3300-6	<u>3317</u> -7, <u>3299</u> -7	<u>3326</u>		NapABCDFGH, NapCE	<u>3298</u> -11	NaTbpa		
Pseudomonas aeruginosa PAO1	4			<u>1782</u> -6	<u>1781</u> -7	<u>1780</u>	NarGHIJ	NapABCDFE			<u>3876, 1784</u>	
Synechocystis sp. PCC 6803	5			<u>2479</u> -10		909				NaTbpa		
Synechococcus sp. WH 8102	5			<u>2468</u> -10		2455					2453, 2454	
Xanthomonas campestris	5			1977-6	1978-7	1979					1976	
Campylobacter jejuni RM1221	6	1501	NrfH					NapAGHBLD				
Haemophilus influenzae R2846	6	89	NrfBCD					NapABCDFGH				
Yersinia pestis KIM	7			<u>160-6</u>	159-7			NapABCDGH	<u>158</u> -12			
Erwinia carotovora	8	<u>3452</u>	NrfBCD	<u>3058</u> -6, 2949-6	<u>2949</u> -7, 3059-7	<u>2948</u>	NarGHIJ	NapABCDFGH		NaTbpa	<u>3225</u>	
Sinorhizobium meliloti 1021	9			5744-6	<u>5745</u> -7	5746		NapABCDFE		NaTbpa	<u>5057</u>	
Pirellula sp. 1	10	<u>6159</u>	NrfH	<u>987</u> -6, <u>215</u> -	<u>2979</u> -7	216, 214				NaTbpa		
Corynebacterium glutamicum	11						NarGHIJ				<u>1143</u>	
Thermotoga maritima MSB8	12			<u>389</u> -8	<u>390</u> -9							
Prochlorococcus marinus str. MIT	12			<u>2230</u> -10					<u>2231</u> -11			
Staphylococcus aureus MRSA252	13			<u>2357</u> -6	<u>2356</u> -7		NarGHIJ				2346	
Porphyromonas gingivalis W83	14	<u>1560</u>	NrfH									
Symbiobacterium thermophilum	15	<u>698</u>	NrfH			<u>1985, 881</u>						
Vibrio cholerae	16							NapABCDFE				
Chromobacterium violaceum ATCC	17					<u>2229</u>	NarGHIJ				<u>3907, 2545,</u> 2228, 2544	
Geobacter metallireducens	18	<u>2519,</u> 2518	NrfH	<u>72-8</u>	<u>71</u> -9		NarGHIJ				<u>2951, 2952</u>	
Geobacter sulfurreducens PCA	19	<u>3133,</u> <u>354</u>	NrfH	<u>1228-8</u>	<u>1229-9</u>							
Shigella dysenteriae M131649	20			2407-6	2408-7		NarGHIJ	NapABCDGH	2409-12		4381	

<u>Functional variants:</u> Bacteria can have different combinations of two general types of NiR and three main types of NaR. At least 20 different functional variants of NiR/NaR patterns have been observed in available bacterial genomes. For example:

#1: as in *E. coli*: assimilatory and respiratory NiRs, respiratory membrane-bound and periplasmic NaRs;

#2. as in *B. subtilis*: assimilatory NiR, assimilatory and respiratory NaRs;

#5: as in most cyanobacteria:only assimilatory NaR and NiR.

Another highly variable component of the pathway is the transport systems for nitrate and nitrite ions.

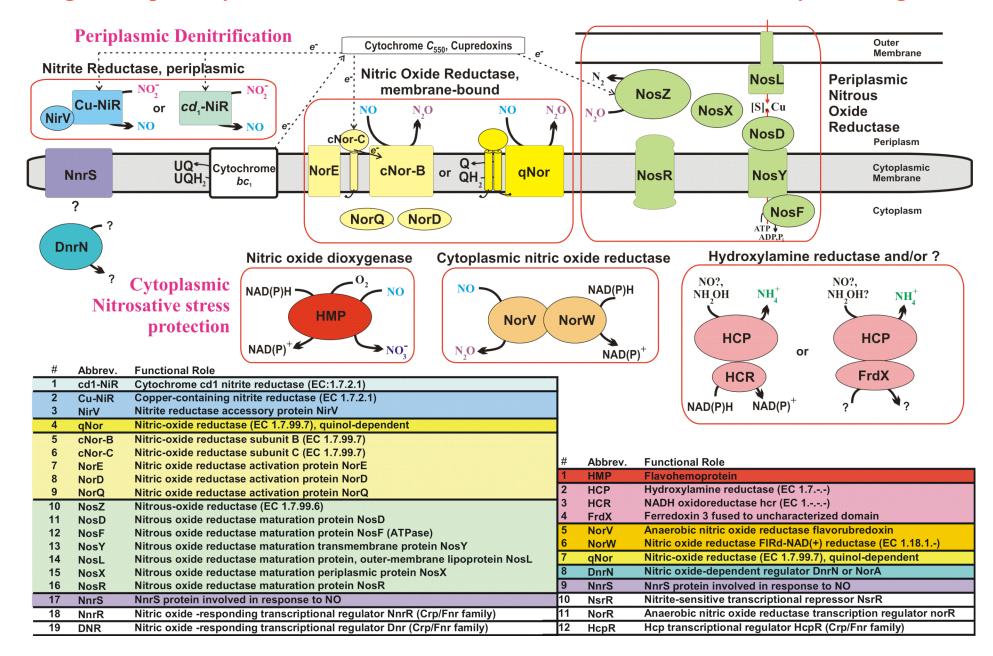
Subsystems: Respiratory Denitrification and and Nitrosative Stress Protection

Denitrification constitutes one of the main branches of the global nitrogen cycle sustained by bacteria [8]. Nitrogen is introduced into the biosphere by fixation of dinitrogen and removed from there again by denitrification. In doing this, denitrification catalyzes successively N-N bond formation in the transformation of its intermediates nitric oxide (NO) and nitrous oxide (N₂O) to the next-lower oxidation state. The bacterial process is nearly exclusively a facultative trait which occurs in periplasm. Its expression is triggered in the cell by the environmental parameters, low oxygen tension, and availability of an N oxide. At the first step, nitrite is formed by one of three different types of nitrate reductases (see Nitrate and Nitrite Ammonification SS), which are shared by the downstream pathways. Next, during denitrification nitrite is reduced to NO by one of the two different types of nitrite reductases (Cu-NiR or cytochrome *cd1*-NiR), then to N₂O by one of the two types of nitric oxide reductase (quinol-dependent qNOR or cytochrome *bc*-type cNOR, [9]), and finally to dinitrogen, using Cu-containing nitrous oxide reductase complex (NOS). Interestingly, the ε -proteobacterium *Wolinella succinogenes* grows anaerobically by respiratory nitrite ammonification but not by denitrification. Nevertheless, it is capable of N₂O reduction to N₂, possesing an active NOS complex [10].

Nitric oxide is a signaling and defense molecule in animals, but bacteria are sensitive to high NO concentrations due to its reactivity and membrane permeability [11]. NO and hydroxylamine (NH₂OH) -- two toxic intermediates in 6-electron reduction of nitrite could be formed during nitrite ammonification [12]. In addition to a classical NO reductase (qNOR or cNOR) occurring in denitrifying species, two other bacterial NO detoxification enzymes have been characterized: an NO reductase (flavorubredoxin NorVW in *Escherichia coli*) and an NO dioxygenase (flavohemoglobin Hmp or Fhp in *E. coli, Bacillus subtilis, Ralstonia eutropha*, and *Pseudomonas* species) [13]. An unusual redox enzyme, called the hybrid cluster protein (HCP) has been extensively studied both in strictly anaerobic and facultative anaerobic bacteria, where it is mostly induced during conditions of nitrite or nitrate stress. *In vitro* studies demonstrated oxygen-sensitive hydroxylamine reductase activity of the *E. coli* HCP protein, suggesting its possible role in detoxification of reactive by-products of nitrite reduction [14].

Recent comparative analysis of NO protection genes and their transcriptional regulatory signals was used to demonstrate considerable interconnection between various regulons of denitrification and NO detoxification and to identify two new members of the nitrosative stress protection pathway, the hypothetical proteins DnrN and NnrS [13].

Fig. 4. Respiratory Denitrification and Nitrosative Stress Protection. Subsystem diagram.



Organism	Variant Code	cd1-Nitrite reductase	Cu-Nitrite reductase	Quinol NO reductase	Cytochro	ome bc-ty	Nitrouse oxide reductase						
		NirS	NirK	qNoR	cNoR-B	cNoR-C	NorE	NorD	NorQ	NosZ	NosD	NosF	NosY
Sinorhizobium meliloti 1021	1		<u>681</u>		<u>694</u>	<u>695</u>	<u>696</u>	<u>692</u>	<u>693</u>	<u>643</u>	644	<u>645</u>	<u>646</u>
Pseudomonas aeruginosa UCBPP-PA	2	<u>5083</u>			5088	<u>5087</u>	<u>5085</u>	5089	5084	<u>976</u>	<u>977</u>	<u>978</u>	<u>979</u>
Neisseria lactamica ST-640	3		563	564						2015	2013	2012	2011
Ralstonia metallidurans	4	3126, 3605		3610						3103	3101	3100	3099
Photobacterium profundum SS9	5				1131	1130	1135	1132	1133	908	909	910	911
Ralstonia eutropha JMP134	6	2868, 2875		2912									
Nitrosomonas europaea ATCC 19718	7		897		1918	1917		1920	1919				
Chromobacterium violaceum	8		2007	3494									
Wolinella succinogenes DSM 1740	9									846, 848	<u>850</u>	855	857
Actinobacillus pleuropneumoniae	10		<u>396</u>										

Fig. 5. Periplasmic respiratory denitrification. Subsystem spreadsheet.

Functional variants:

#1 and 2: two variants of a complete denitrification pathway from nitrite to N_2 , which use different types of nitrite reductase. #3, 4, 6, 7, 8: five variants of the denitrification pathway from nitrite to N_2O , which use different types of nitrite and NO reductases. #10: short denitrification pathway from nitrite to NO. #9: solely N₂O reduction pathway. #5: solely NO and N₂O reduction pathway.

Fig. 6. Cytoplasmic nitrosative stress protection. Subsystem spreadsheet.

					-				
Organism	Variant Code	NO dioxygenase	NH2OH reductase	Anaerobic NO reductase		NO reductase	N	s	
		HMP	HCP	NorV	NorW	qNOR	DnrN	NnrS	
Escherichia coli K12	1N	2522	859	2665	2666		4119		
Vibrio vulnificus CMCP6	1S	1196	2634	4202	4203			1625	
Yersinia pestis KIM	2N	1295	2771				641		
Vibrio parahaemolyticus	2S	2809	<u>1189</u>					2390	
Bacillus subtilis	3	1306							
Staphylococcus aureus NCTC 8325	3N	216					235		
Vibrio cholerae O1	35	<u>2924</u>						2302	
Microbulbifer degradans 2-40	3NS	<u>2172</u>					461	<u>2616</u>	
Desulfovibrio desulfuricans G20	4		<u>924</u>						
Clostridium perfringens str. 13	4N		<u>2647</u>				838		
Rhodospirillum rubrum	4S		<u>407</u>					<u>3252</u>	
Shewanella oneidensis MR-1	4NS		<u>1266</u>				3904	<u>2551</u>	
Chromobacterium violaceum	5S	<u>3488</u>				<u>3494</u>		2711	
Ralstonia eutropha JMP134	5NS	<u>5127</u>				<u>2912</u>	<u>2913</u>	2856	
Mannheimia succiniciproducens	6N		<u>278</u>			<u>280, 281</u>	<u>1164</u>		
Synechocystis sp. PCC 6803	7					<u>3084</u>			
Neisseria meningitidis Z2491	7NS					<u>1740</u>	<u>1436</u>	<u>1890</u>	
Haemophilus influenzae R2866	8N						<u>1056</u>		
Rhodopseudomonas palustris CGA009	8S							<u>1478</u>	

<u>Functional variants</u>: Bacteria can have eight different combinations of four known nitrosative stress protection systems (HMP, HCP, qNor and NorVW). These basic variants are further subdivided into sub-variants (marked by additional letters N and S) depending on the presence of the two hypothetical genes *dnrN* and/or *nnrS*, which are often co-regulated or co-localized with nitrosative stress protection and denitrification genes, and thus should play an important functional role in these processes.

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