Subsystem: Thiamin biosynthesis

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Thiamin pyrophosphate (vitamin B1) is an essential cofactor for several important enzymes of the carbohydrate metabolism. Many microorganisms, as well as plants and fungi synthesize thiamin, but it is not produced by vertebrates. Thiamin monophosphate is formed by coupling of two independently synthesized moieties, hydroxymethylthiamin-PP (HMP-PP) and hydroxyethylthiazole-P (HET-P).

The HET moiety is biosynthesized in *Bacillus subtilis* and most other bacteria from DXP, Glycine, and cysteine in a complex oxidative condensation reaction [1]. This reaction requires five different proteins, ThiO, ThiG, ThiS, ThiF, and a cysteine desulfurase. Glycine oxidase ThiO catalyzes the oxidation of glycine to the corresponding glycine imine. Sulfur carrier protein adenylyl transferase ThiF catalyzes the adenylation of the carboxy-terminus of the sulfur carrier protein ThiS, and cysteine desulfurase catalyzes the transfer of sulfur from cysteine to the ThiS-acyl adenylate to give ThiS-thiocarboxylate. ThiG is the thiazole synthase and catalyzes formation of the thiazole from dehydroglycine, DXP, and ThiS-thiocarboxylate [2, 3, 4]. The thiazole moiety of thiamin in *E. coli* is derived from Tyrosine, cysteine, and DXP using another enzyme (ThiH) of yet unknown function instead of ThiO [5]. In contrast, the THI4 protein family has been shown to be involved in the thiazole synthesis in some eukaryotes [7]. The HET kinase ThiM is involved in the salvage of thiazole from the culture medium.

The conversion of 5-aminoimidazole ribonucleotide (AIR) into HMP is a fascinating reaction of the thiamin biosynthetic pathway in bacteria and is probably the most complex unresolved rearrangement in primary metabolism. The *thiC* gene complements all HMP requiring mutants in *E.coli*, and *B.subtilis* [6]. In yeast and plants, the pyrimidine moiety of thiamin is synthesized using a distinct gene (THI5 in yeasts), and the initial substrates appear to be histidine and pyridoxol-P [8, 9]. HMP-P is phosphorylated by the bifunctional HMP kinase/HMP-P kinase ThiD.

The ThiE protein catalyzes the formation of thiamin-P via coupling of HMP-PP and HET-P moieties. The ThiN protein recently identified in some ThiE-lacking archaea and in *T. maritima* [10] could complement *E.coli thiE* mutant strain and thus presents a case of non-orthologous gene displacement of the thiamin-P synthase ThiE [11]. The thiamin-phosphate phosphatase activity is present in yeast but corresponding gene has not yet been characterized. Bacteria synthesize TPP via single phosphorylation of TP using the ThiL kinase, whereas eukaryotes use distinct pathway to form an active coenzyme TPP: hydrolysis of TP to free thiamin is followed by pyrophosphorylation [8].

Search for thiamin-specific regulatory elements (*THI* riboswitches) and analysis of operon structures identified a large number of new candidate thiamin-regulated genes, mostly transporters, in various bacterial genomes [10]. In particular, the thiamin transporter function was assigned to *yuaJ* in the *Bacillus/Clostridium* group and *thiT* in Archaea. Also previously unknown HMP and HET transporter functions were tentatively assigned to several genes including *thiXYZ* and *ykoEDC* (ABC-type HMP transporters), *cytX* (HMP permease), *thiU* and *thiW* (HET permeases). Identification of the predicted uptake systems for thiamin precursors, HET and HMP, allowed to reconstruct the thiamin pathways in organisms that are unable to synthesize HET and HMP *de novo* and are forced to uptake them via specific transport system [10].

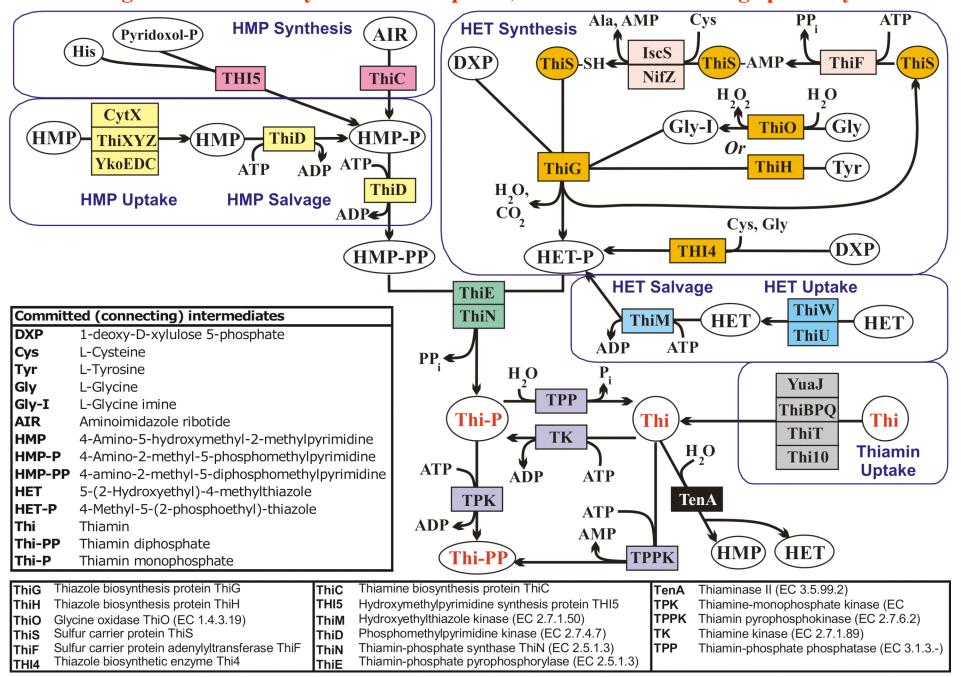


Fig. 1. Thiamin biosynthesis and uptake; HET and HMP salvage pathways

		HMP synthesis	HET synthesis		Thi-P synthesis	HMP salvage		HET salvage		Thiamin Uptake	
Organism		ThiC-1; THI5-35		ThiH-3; ThiO-4	THI4	ThiE-11; ThiN-9	ThiD	HMP Uptake: CytX-21; ThiY-14,ThiX-15,ThiZ-16; YkoE-25,YkoD-26,YkoC-27		HET Uptake: ThiW-22; ThiU-23	ThiB-18, ThiP-19, ThiQ-20; YuaJ-17, ThiT-24, THI10-36
Shewanella oneidensis MR-1	1	2216-1	2212	2211-3		2215-11	2215				
Escherichia coli K12	11	3907-1	3903	3902-3		3906-11	2078		2079		69-18, 68-19, 67-20
Mycobacterium tuberculosis CDC1551	2	438-1	431	429-4		428-11	437				
Streptomyces coelicolor A3	22	3885-1	2075	2073-4		2070-11	5516				5595-18, 5596-19, 5597-20
Thermotoga maritima MSB8	3	<u>781</u> -1			780	783-9	783	478-14, 479-15, 477-16			
Pyrobaculum aerophilum str. IM2 [A]	33	229-1			122	<u>1538</u> -9; <u>1770</u> -9	<u>1538,</u> 1770				227-18, 226-19, 225-20; 123-24
Neurospora crassa [E]	4	9970-35			3898	184 -11	6488		184		
Saccharomyces cerevisiae [E]	44	<u>1690</u> -35; <u>3168</u> -35; <u>4461</u> -35			<u>2183</u>	<u>5452</u> -11	<u>4958,</u> 5407, 5764		<u>5452</u>		<u>3779</u> -36, <u>5080</u> -36, <u>5194</u> -36
Methanococcus maripaludis S2 [A]	5	<u>187</u> -1			<u>1352</u>	<u>1639</u> -9; <u>1139</u> -11	<u>1639</u>		<u>1138</u>		
Pyrococcus abyssi GE5 [A]	55	683-1			827	<u>1140</u> -9; <u>1413</u> -9; <u>1141</u> -11	<u>1140,</u> 1413	1143-21			836-18, 834-19, 835-20
Archaeoglobus fulgidus DSM 4304 [A]	6	2396-1			695	2059-11	2194		2060		
Streptococcus pneumoniae R6	7					<u>630</u> -11; <u>637</u> -11	<u>1449,</u> 638	<u>2000</u> -14, <u>2001</u> -15, <u>1999</u> -16; <u>631</u> -25, <u>632</u> -26, <u>633</u> -27		<u>635</u> -22	
Haemophilus influenzae R2866	77					<u>1610</u> -11	<u>1609</u>	<u>1550</u> -14 <u>, 1549</u> -15 <u>, 1548</u> -16	1608	<u>1611</u> -23	472-18, 473-19, 474-20
Bradyrhizobium japonicum USDA 110	8	<u>6659</u> -1	<u>6657</u>	<u>6655</u> -4		<u>6658</u> -11	<u>5905</u>	2837-14, 2836-15, 2838-16			
Bacillus subtilis subsp. subtilis str. 168	88 9	<u>878</u> -1	<u>1170</u>	<u>1168</u> -4 46-4		<u>1167</u> -11; <u>3834</u> -11	<u>1172,</u> <u>3807</u> 45		<u>3835</u>		<u>3102</u> -17
Silicibacter pomeroyi DSS-3	-		48			<u>49</u> -11		51-14, 54-15			004 40 002 40 000 00
Brucella melitensis 16M	99		1734	<u>1732</u> -4		<u>1735</u> -11	1731	<u>1736</u> -14, <u>1738</u> -15			284-18, 283-19, 282-20
Streptococcus pyogenes M5	10 10						<u>161</u> 114				<u>1584</u> -17 143-18, 142-19, 141-20
Treponema pallidum subsp. Pallidum	10						114				140-10, 142-13, 141-20

Fig. 2. Thiamin biosynthesis. Subsystem spreadsheet.

Functional variants:

#1: HMP synthesis (ThiC-1), HET synthesis (ThiG,ThiH-3), thiamin synthase (ThiE-11);

#2. HMP synthesis (ThiC-1), HET synthesis (ThiG,ThiO-4), thiamin synthase (ThiE-11);

#3: HMP synthesis (ThiC-1), HET synthesis (THI4), thiamin synthase (ThiN-9);

#4: HMP synthesis (THI5-35), HET synthesis (THI4), thiamin synthase (ThiN-9);

#5: HMP synthesis (ThiC-1), HET synthesis (THI4), thiamin synthases (ThiE-11 and ThiN-9);

#6: HMP synthesis (ThiC-1), HET synthesis (THI4), thiamin synthase (ThiE-9);

#7: HMP and HET salvage, thiamin synthase (ThiE-11);

#8: HMP synthesis (ThiC-1), HET synthesis (ThiG,ThiO-4), thiamin synthase (ThiE-11), HMP salvage;

#9: HMP salvage, HET synthesis (ThiG,ThiO-4), thiamin synthase (ThiE-11);

#10: only Thiamin Uptake;

*Variants #11,22,33,44,55,77,88,99 are the same as #1,2,3,4,5,7,8,9, respectively, but also include additional genes for Thiamin Uptake. Analysis of regulatory elements. Predicted thiamin-regulated transporter genes [10]

1. YuaJ: predicted Thiamin Transporter (possibly H⁺-dependent)

- Found only in bacteria from the *Bacillus/Clostridium* group
- All these species lack the known thiamin ABC transporter ThiBPQ;
- Occurs in genomes without thiamin biosynthetic pathway (Streptococcus spp.);
- Has 6 predicted transmembrane segments;
- Regulated by thiamin riboswitches;
- Bacillus cereus is able to uptake thiamin in a proton-dependent manner [12].

2. ThiXYZ and YkoEDC: predicted ATP-dependent HMP Transporters

- Found in some Proteobacteria and Firmicutes;
- Not found in genomes without thiamin biosynthetic pathway;
- Always co-occur with the *thiD* and *thiE* genes;
- Present in Pasteurellae, Brucella and Gram-positive cocci, lacking HMP synthase ThiC;
- Regulated by thiamin riboswitches;
- The substrate-binding component **ThiY** is homologous to the HMP synthase **THI5** from Yeast and has an N-terminal signal peptide for possible membrane binding.

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